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SYNTHESIS AND BIOLOGICAL EVALUATION OF 1, 3, 4 THIADIAZOLE DERIVATIVE ON SOME PARAMETERS OF IMMUNITY AND LIVER ENZYMES

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ABSTRACT

In the present study, a series of five members heterocyclic where synthesized by the reaction between isoniazid and various substituted isothiocyanates. The newly synthesized compounds where characterized by IR and $^1\text{H-NMR}$ spectral data. The effect of Thiadiazole derivative (2b) in monocyte and Lymphocyte in the differential count is 20349 and 11415 cells/cu.mm, respectively. Prepared compounds (2a&2b) where not effective against differential count of Eosinophil. In WBCs differential count that significant immune effects were occurring in compound (2b) more than in compound (2a). Liver enzymes, glutamic oxaloacetic acid transaminase (GOT) and glutamic pyruvic acid transaminase (GPT) were chosen to assess liver function. In comparison with control, the administration of Thiadiazole derivatives (2a and 2b) significantly declined the activity of GOT, GPT and Urea. These results indicate that prepared compound effectively increases of immune system in the animals.

Keywords: 1,3,4-thiadiazoles derivatives, spectral data, biological evaluation, packed cell volume.

1. INTRODUCTION

In recent years 1, 3, 4-Thiadiazole derivatives have received significant attention and have been increasingly investigated due to their diverse range of biological properties. They exhibit for example, antimicrobial^{1,2}, anti-microbial³, anticancer⁴, anti-inflammatory^{5,6}, carbonic anhydrase inhibiting effect⁷, anti-anxiety, anti-depressant⁸, anti-oxidant properties⁹.

1,3,4-Thiadiazole exhibit diverse biological activities, possibly due the present of $=\text{N-C-S}$ moiety¹⁰. 1, 3,4-Thiadiazole are very interesting compounds due to their important applications in many pharmaceutical biological and analytical fields^{11,12}. The therapeutic importance of these rings prompted us to develop selective molecules in which a substituted could be arranged in a pharmacological activity. Derivatives of these nuclei are synthesized from

substituted thiosemicarbazides, obtained by reaction isonized and different substituted isothiocyanates. There are five distinctly different kinds of board cells (WBCs), neutrophils, monocytes, Lymphocytes, eosinophils and basophils, some have ability to change with needs and situations in the body. So, for example, there are different monocytes found in different tissues, and different types of Lymphocytes with different rules in fighting infections. These cells can leave the bloodstream sliding out through the vessel walls and attacking invaders at the site of an infection¹³. Liver is in the central organ of metabolism and act as an organ of storage. Many potentially toxic substances are metabolized by cells, metabolic action by the hepatic parenchyma cells have been regarded as an important defense system against

toxicants and the transformation involved have been referred to as detoxification.

The great susceptibility of liver to damage by chemical agent is presumably a consequence of its primary role in metabolism of foreign substances. The role of liver in metabolic conversion is due to its susceptibility to chemical injury¹⁴. Liver enzyme such as glutamic oxaloacetic transaminase (GOT), glutamic pyruvic transaminase (GPT), and urea are considered to be biochemical makers for assessing liver function.

2. MATERIALS AND METHODS

Melting points were determined by open capillary tube method and are un-corrected. Purity of the compounds was checked in thin layer chromatography (TLC) plates (silica gel G) in the solvent system toluene: ethyl acetate: formic acid (5:4:1, v,v,v) and benzene: acetone (8:2,v,v). The IR spectra were obtained on a Perkin-Elmer 1720 FT-IR spectrometer (KBr pellets). The H-NMR Spectra were obtained on a Bruker Ac 300MHz spectrometer in (DMSO-D6) using TMS as an internal standard and mass spectra under electron impact conditions (EI)

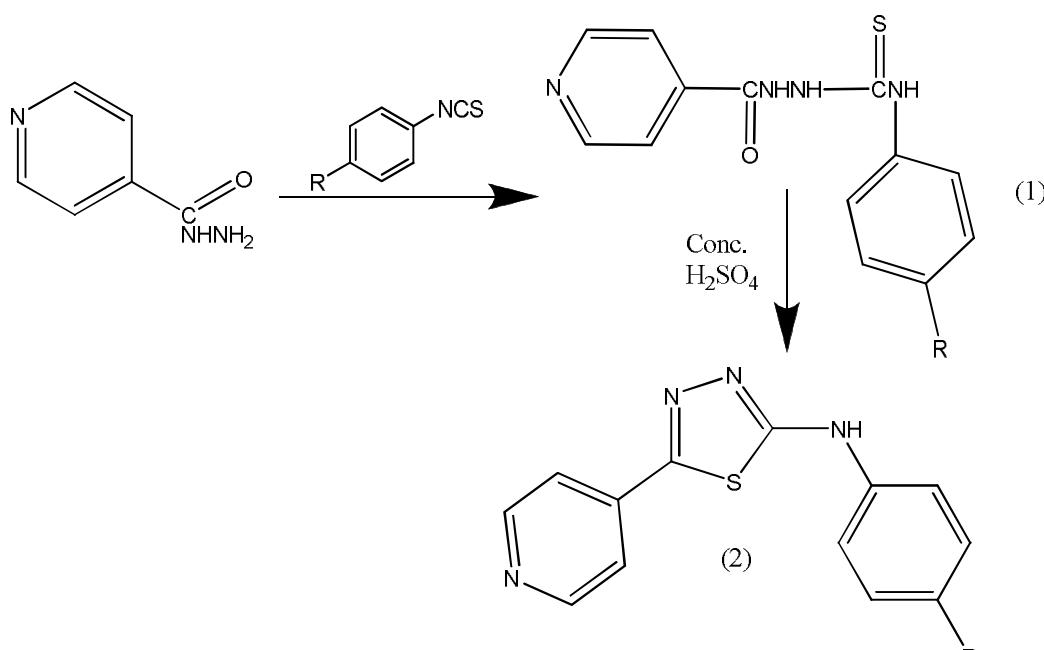
were recorded at 70 ev ionizing voltage with AG prospect instrument and are presented as m/z.

2.1. Method synthesis of thiosemicarbazide

Substituted phenyl thiosemicarbazide (1) was synthesized by refluxing isonized (0.02 moles) with substituted phenol isothiocyanates (0.02 moles) in 15ml ethanol on a boiling water bath for 6hr. after completion of reaction, the reaction mixture was concentrated and kept overnight at room temperature. The needle shaped crystals of thiosemicarbazides.

2.2. Method synthesis of 2-(substituted phenyl)-amino-5-(4-pyridyl)-4H-1,3,4-Thiadiazole²

2-(substituted phenyl)-amino-5-(4-pyridyl)-4H-1,3,4-thiadiazole(2) were synthesized by cyclization of substituted phenyl thiosemicarbazide of isonized (0.002 moles) with sulfuric acid at 0-5°C. After completion of reaction, the mixture was poured into crushed ice, the solid separated was filtered, washed with water and re-crystallized from methanol yielded the pure compound.



R= -OCH₃, - Cl

Scheme 1: Synthetic Pathways for the Preparation of 1, 3, 4-Thiadiazole Derivatives

Table 1: Physical Constant of Newly Synthesized Thiadiazole Derivatives

Compound	R Group	Yield(%)	m.p. (°C)	Mol. Formula	Mol. Wt.
2a	-Cl	70	152-54	C ₁₃ H ₉ CIN ₄ S	288.75
2b	-OCH ₃	81	168-70	C ₁₄ H ₁₂ N ₄ OS	284.33

2.3. Blood Film Methods

This method was done according to Catalovo A.¹⁵.

2.4. Differential Count of Leukocytes

1. A small drops of heparinized blood which drawn from mouse was put on the end of clean and dry slide. A pusher slide was place at an angle of 30 to 45°C to the slide and then moved back to make contact with the drop. The forward movement of the pusher spreads the blood on the slide.
2. The blood film was allowed to dry in air.
3. The slides were completely covered with Leishman stains, after 3min. the slides were washed gently and then examined under light microscope and by applying the following equation:

$$\text{No. of cells/ (cells/mm}^3 \text{ blood)} = (\text{total no. of leukocytes \%}) / 100$$

2.5. Total Count of Leukocytes

1. The blood was taken by heart puncher and put into heperinized tube.
2. A dilution solution (190μL) was pipette into test tube.

3. The heperinized Blood (10μL) was pipette and mixed well with diluting fluid for at least 2minutes.
4. The hemocytometer was sited up with its cover glass in position and by a pasture pipette; both sides of the hemocytometer were filled with the diluted blood.
5. The cells were allowed for two minutes to be settled.
6. The cells were count in the four large squares on both sides of chamber using the 40X objectives and subdued light.
7. The WBCs were calculated on the bases of cells counted area, and the dilution.

No. of cells/ (cells/nm³ blood) = No. of cells in four square X correct volume correct dilution /4
According to Creskoff.¹⁶, blood was collected from the mice by heart puncture. The serum was separated by centrifuge at 2000 rpm for 10 min. then; the serum was taken and treated as follows: two test tubes were used for each sample; the first one contained the blank reagent and second contains the sample. These samples were tested as in the following:

Table 2: Method of Total Count of Leukocytes

	GPT	GOT
Reagent 1	1ml	-
Reagent 1	-	1ml
Serum	0.2ml	0.2
Mix and incubate at 37°C	1 hour	30 min.
Reagent 3	1ml	1ml
Mix let stand for 20min at room temp.	-	-
NaOH 0.4N	10ml	10ml

Mix wait 5min. measure under condition identical to those used for the standard curve.

Wave length: 505nm (490-520nm)

Activities for these two enzymes in the serum were estimated from the activity table attached with kit of each enzyme.

2.6. Statistical Analysis

Statistical analysis was performed to compare two different groups by using ANOVA-test. Statistical significance was determined at (P<0.05)¹⁷.

3. RESULTS AND DISCUSSION

A series of 2-(substituted phenyl)amino-5-(4-pyridyl)-4H-1,3,4-thiadiazole was prepared from

ionized and substituted phenyl isothiocyanates derived thiosemicarbazides (scheme 1). The structure of newly synthesized compounds was confirmed by spectral and analytical data. In general, the IR spectra of newly synthesized compounds revealed NH, C=N, N-N, C-S-C peaks near 3390, 1620, 1065 and 660cm⁻¹, respectively. In the ¹H-NMR spectra, signal of respectively protons of newly synthesized compounds showed the peaks for –OCH₃, NH & aromatic protons near δ3.7, 7.5 and 6.8-8.5, respectively.

The general mass fragmentation pattern for the compounds showed the m/z peaks. Both analytical and spectral data (IR, ¹H-NMR and mass) of all the synthesized compounds were in full agreement with proposed structures. Physical data of all the synthesized compounds are presented in Table 1.

3.1. Spectral Data

3.1.1. Compound of 2-(4-chlorophenyl)-

amino-5-(4-pyridyl)-4H-1,3,4-thiadiazole(2a)

IR(KBr,cm⁻¹):3390(NH), 1620(C=N), 1060(N-N)

660 (C-S-C); (400 MHz, DMSO-d6): 8 7.4-7.7

(d,2H,Ar, J=8.56Hz), 8.4-8.9 (d,2H, pyridine,

J=8.71Hz), 8.0 (S, 1H, NH), MS (m/z):289

(M⁺+1).

3.1.2. Compound of 2-(4-methoxyphenyl)-amino-5-(4-pyridyl)-4H-1,3,4-thiadiazole(2b)

IR(KBr,cm⁻¹):3070(NH), 1610(C=N), 1003(N-N)

700 (C-S-C); (400 MHz, DMSO-d6): 8 3.7(S, 3H,

OCH₃), 6.8-7.5 (d,2H,Ar, J=8.25Hz), 8.2-8.8

(d,2H, pyridine, J=5.50Hz), 7.6 (S, 1H, NH), MS

(m/z):284 (M⁺).

3.2. Biological Evaluation

Its aimed to study effects 1,3,4-Thiadiazole derivatives on blood film (total and differential) (WBCs) and liver enzymes. Animals were divided into three groups of 6 animals each as follows: none treated mice (control group), mice were injected with compound (2a), and mice were injected with compound (2b).

3.2.1. White Blood cells (WBCs)

The compound (2b) was found to be the most active in Lymphocyte and monocyte in differential count 11415 and 20349 cells/cu.mm, respectively, while compound (2a) showed little activity against all types of white blood cells (WBCs) by compared with control. These prepared compounds (2a and 2b) were not effective against the count of Eosinophil, as shown in Table 2. and figure 1.

Table 3: The Effect of 1,3,4- Thiadiazole Derivatives on Differential and Total Counts of Blood

Groups	Total WBCs (m±SE)%	Differential count (m±SE)				
		No. of Lymphocyte cells/cu. mm blood	No. of Neutrophil cells/cu. mm blood	No. of monocyte cells/cu. mm blood	No. of Eosinophil cells/cu. mm blood	No. of Basophil cells/cu. mm blood
Control	A 100.63±11.41	A 6100±98.4	A 2410±121.6	A 1629±200	A 71.0±21.0	A 0.00±0.00
Treated with 2a	B 153.5±14.1	B 10122±100.5	B 3612±201.3	B 2323±182.0	B 0.00±0.00	A 0.00±0.00
Treated with 2b	C 171.0±16.2	B 11415±321.4	C 4082±141.6	B 20349±124.6	B 0.00±0.00	B 526.3±74.8

* Differences A, B and C are significant (P<0.05) to compression column

3.2.2 Packed Cell Volume and Hemoglobin

These two compounds (2a and 2b) of 1,3,4-thiadiazole derivatives that were prepared in theses research showed high effect in total and differential count of white blood cells (WBCs), percentage of packed cell volume (P.C.V %), and hemoglobin percentage (H.B %). As shown in Table 3.

Table 4: The Effect of 1,3,4- Thiadiazole Derivatives On Packed Cell Volume And Hemoglobin

Groups	P.C.V % (mean ± SE)%	Hb % (mean ± SE)%
Control	A 28.56±3.20	A 9.64±1.45
Treated with 2a	AB 31.27±2.98	B 11.33±2.63
Treated with 2b	B 36.45±3.61	C 13.51±1.94

Differences A, B and C are significant (P<0.05) to compression column

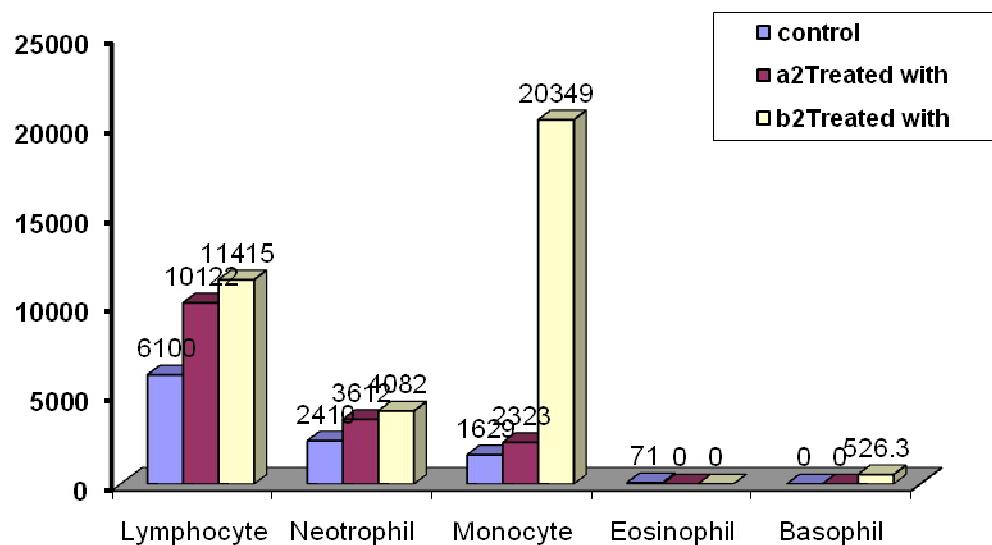


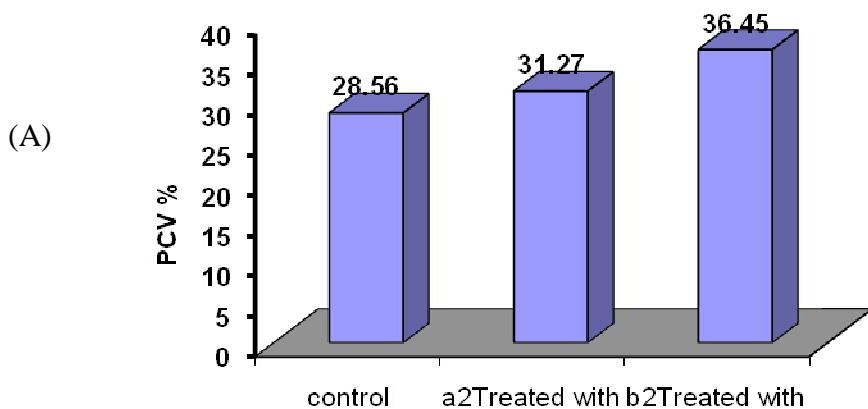
Fig. 1: The relationship between treated of Thiadiazole derivatives (2a,2b) and differential count of WBCs

Immune mechanisms affected by Thiadiazole derivatives, in addition to blood film dependent immunity, include reduced production of complement by the liver and decreased phagocytosis by neutrophils and microphage¹⁹. WBCs generally fall into distinct subtypes:

- 1) Polymorph-nuclear leukocytes of granulocytic lineages, including neutrophils, eosinophils and basophils.

- 2) Lymphomononuclear cells such as Lymphocytic and monocytic²⁰.

The compound (2b) emerged as the most active against the packed cell volume (PCV %) at (mean \pm SE) 36.45 ± 3.61 , and hemoglobin (Hb %) 13.51 ± 1.94 , as shown in figure 2.



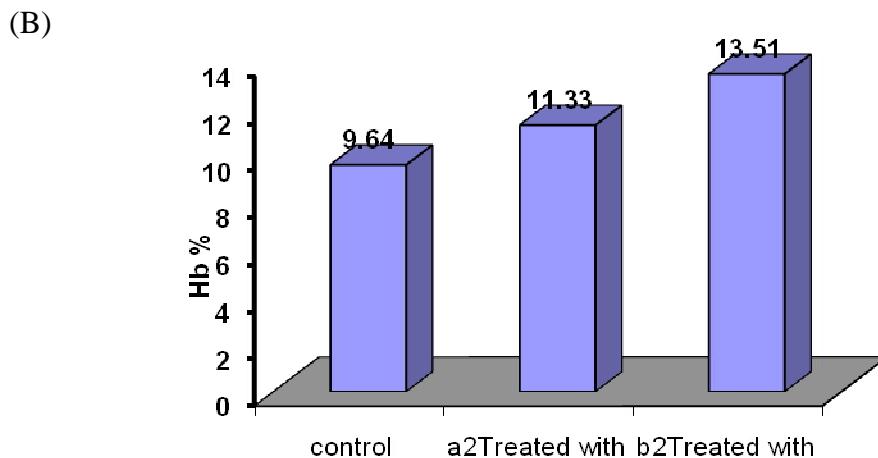


Fig. 2: (A) The Percentage of Packed Cell Volume. (B) Relationship between Treated of Thiadiazole Derivatives and Percentage of hemoglobin.

Hemoglobin gives red blood cells their color. Hemoglobin carries from the lungs to the tissues and takes carbon dioxide from the tissues to the lungs²¹.

3.2.3. Liver Enzymes

The present study showed that the exposure of 1,3,4-Thiadiazole derivatives caused decreasing the activity of glutamic oxaloacetic acid transaminase (GOT), glutamic pyruvic acid transaminase (GPT), and urea.

The compound (2b) was found to be the most active against GOT, GPT and urea, while compound (2a) showed moderate activity against these tests.

A significant decrease in the serum GOT (188.7 ± 12.2) & GPT (58.7 ± 4.81) IU/ml levels were seen in the compound (2b) treated mice, by compared with control, the results of such studies are given in Table 4.

Table 5: Effects of 1, 3,4- Thiadiazole Derivatives On Activity of liver function represent in glutamic oxaloacetic acid transaminase (GOT), glutamic pyruvic acid transaminase (GPT), and urea

Groups	GOT IU/ml (m ± SE)%	GPT IU/ml (m ± SE)%	Urea mg/dl (m ± SE)%
Control	A 210.4 ± 16.8	A 68.5 ± 10.6	A 15.6 ± 4.30
A Treated with 2a	B 198.6 ± 14.5	B 62.3 ± 6.40	B 12.2 ± 3.8
B Treated with 2b	C 188.7 ± 12.2	B 58.7 ± 4.81	B 10.4 ± 2.83

Differences A, B and C are significant ($P < 0.05$) to compression column

The marked elevation of urea level in the serum of mice were significantly decreases in the Thiadiazole, the GOT & GPT levels respectively dropped from 210.8 ± 16.8 IU/ml to 198 ± 14.5 IU/ml regarding compound (2a) and from 210.4 ± 16.8 IU/ml to 188.7 ± 12.2 IU/ml regarding compound (2b), as shown in figure 3.

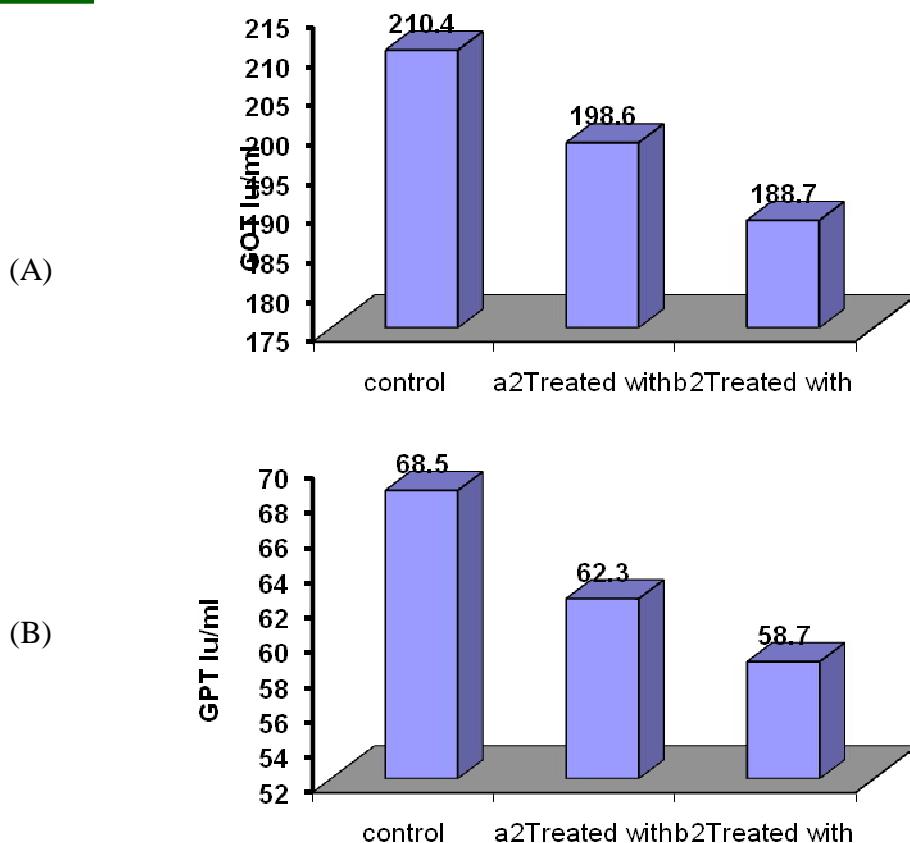


Fig. 3: Effect of prepared compound (2a and 2b) in liver enzymes, (a) glutamic oxaloacetic acid transaminase (GOT), and (b) glutamic pyruvic acid transaminase (GPT).

Liver enzymes, GOT & GPT were chosen to assess liver function. The enzyme is absorbed by the organism and passes through the blood to fulfill a systemic activity. Thus, it inhibits the production of prostaglandins which provoke inflammations. The decrease in inflammatory phenomenon results in the retrogression of liver deterioration. The results establish the decrease in those two enzymes and urea. On the whole, the use of Thiadiazole derivatives made the GPT & GOT rates increase the Liver function.

CONCLUSIONS

A series of 1,3,4-Thiadiazole derivatives were synthesized and their structures were elucidated by spectral data. The biochemical revealed that the 1,3,4-Thiadiazole caused activator effects on GOT, GPT, and urea enzymes activities, and increase the differential count of WBCs, PCV, and Hemoglobin percentage. The compound (2b) was found to be the most active against GOT, GPT and urea, while compound (2a) showed moderate activity against these tests.

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REFERENCES

- Demirbas N, Karaoglu SA and Sancak K. Synthesis and antimicrobial activities of some new 1- (5-phenylamino-[1,3,4] thiadiazole-2-yl)methyl-5-oxo-[1,2,4]triazole and 1-(4-phenyl-5-thioxo-[1,2,4]triazole-3-yl)methyl-5-oxo-[1,2,4]triazole derivatives. Eur J Med Chem. 2004;39:793-804.
- Desai K and Daxi A. Studies on 2-azetidinone: part VI synthesis and antimicrobial activity of 5-(2',4'-dichlorophenoxyethyl)-2-(4",aryl-3"-chloro-2"-azetidinone-1"-yl)-1,3,4-thiadiazole. Indian J Pharm Sci. 1992;54:183-188.

3. Matysiak J, Nasulewicz A, Pelczynska M, Switalska M, Jaroszewicz I and Opolski A. Synthesis and anti-proliferative activity of some 5-substituted-2-(2,4-dihydroxyphenyl)-1,3,4-thiadiazoles. *Eur J Med Chem.* 2006;41:475-482.
4. Azam M, Kumar B, Shalinis S, Surech B, Reddy K and Reddy D. Synthesis and biological screening of 5-[(4,6-disubstituted pyrimidine-2-yl)thio]methyl]-N-phenyl-1,3,4-thiadiazole-2-amines. *Indian J Pharm Sci.* 2008;70:672-677.
5. Foroumadi A, Emani S, Hassan Zadeh A, Rajee M, Sokhanvar K and Moshafi H. Synthesis and antibacterial activity of N-(S-bezylthio-1,3,4-thiadiazole-2-yl) piperazinyl quinolone. *Bio Org Med Chem Lett.* 2005;15:4488-4492.
6. Chou Y, Lais Y, Pan L, Jow M, Chen W and Guh H. Investigation of anticancer mechanisms of thiadiazole-based compound in human non-small cell lung cancer A 549 cells. *Biochemist. Pharmacology.* 2003;66:115-124.
7. Nadeem S, Priya A, Waquar A, Pandeya S and Shamsher M. Thiadiazoles: Progress report On Biological Activities. *J Chem Pharm Res.* 2009;1(1):19-30.
8. Cleici F, Pocar D, Guido M, Loch A, Perlini V and Brufani M. Synthesis of 2-amino-5-sulfanyl-1,3,4-thiadiazole derivatives and evaluation of their antidepressant and anxiolytic activity. *J Med Chem.* 2001;44(6):931-936.
9. Martinez A, Alonso D, Castro A, Aran J, Cardelus I and Banos E. Synthesis and Potential muscarinic receptor binding and antioxidant properties of 3-(thiadiazolyl) pyridine 1-oxide compounds. *Arch Pharm.* 1999;332:191-194.
10. Oruc E, Rollas S, Kandemirli F, Shvets N and Dimoglo A. 1,3,4-thiadiazole derivatives, synthesis, structure elucidation and structure antituberculosis activity relationship investigation. *J Med Chem.* 2004;47:6760-6767.
11. Katritzky A, Rees CW and Potts KT (eds.), *Comprehensive Heterocyclic Chemistry.* Oxford-VCH, 6(1982).
12. Ahmed M, Jahan J and Banco S. A simple spectrophotometric Methods for the determination of copper in Industrial, Environmental, Biological and Soil, samples using 2,5-dimercapto 1,3,4-thiadiazole. *J anal Sci.* 2002;18:805-810.
13. Denis M, Michael J, Richard A, Sabine F, Rene E, Josephs S and Ming W. Immunotoxicity of Aflatoxin B1 in rats: effects on lymphocytes and the inflammatory response in a chronic intermittent dosing study. *J Toxicol Sci.* 2003;73:362-377.
14. Guntupalli M, Chandana V, Palpu P and Annie Shirwaikar I. Hepatoprotective effect of rubiadin, a major constituent of Rubiacordifolia Linn. *J. Ethnopharmacol.* 2006;103:484-490.
15. Catalovo A. White Blood Cell count With Differential, in George-Gay, B. and Chernecky (eds). *Clinical medical nursing sounders.* 2002:282-290.
16. Wood RF. Statistical Methods for the Analysis of biomedical data, Probability and Mathematical Statistics, Whily, Chichrster. 1983:315-316.
17. Liu BH, Yu FY, Cham MH and Yang YL. The effect of mycotoxins, fuonisin and aflatoxin B1, on primary swin alveolar macrophages. *Toxicol Appl Pharmacol.* 2002;108:197-204.
18. Cusumano V, Costa GB, Trifiletti R, Merendino R and Mancuso G. Functional important of rat kupffer cells included by aflatoxin B1and its metabolites. *FEMS Immunol Med Microbial.* 1995;19910:151-155.
19. Dugyala RR and Sharma RP. The effect of aflatoxine on cytokine mRNA and corresponding protein levels in peritoneal macrophages and splenic lymphocytes. *Int J Immunol pharmacol.* 1996;18:599-608.
20. Batey RG and wang J. Molecular Pathogenesis of T-lymphocyte induced liver injury in alcoholic heptatites. *Front Bio Sci.* 2002;7:1662-1675.
21. Dennis M, Michael J, Richard A, Sabine F, Rene E, Joseph S, Alan W and Ming W. Immunotoxicity of aflatoxin B1 in rats: effects on Lymphocytes and the inflammatory in a chronic intermittent dosing study. *J Toxic Sci.* 2003;73:362-377.